

Enzyme-Activated Irreversible Inhibitors of Monoamine Oxidase: Phenylallylamine Structure-Activity Relationships

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Seventeen 2-aryl-3-haloallylamine derivatives were prepared and evaluated as inhibitors of monoamine oxidase (MAO, EC 1.4.3.4). The synthesis of these compounds was achieved from either α -methylstyrene or ring-substituted phenylacetic acid derivatives. With one exception, these 2-arylallylamines were found to be enzyme-activated, irreversible inhibitors of MAO. The most potent inhibitors were ring-substituted derivatives of (*E*)-2-phenyl-3-fluoroallylamine with IC_{50} values ranging from 10^{-6} to 10^{-8} M. Selectivity for the A and B form of MAO was found to depend on the nature of aromatic ring substitution. In general, hydroxyl substitution favored the inactivation of the A form of MAO, while very selective B inhibitors were obtained when the aromatic ring was substituted with a 4-methoxy group. (*E*)-2-(4-Methoxyphenyl)-3-fluoroallylamine and (*E*)-2-(3,4-dimethoxyphenyl)-3-fluoroallylamine proved to be in vitro as selective for the B form of MAO as deprenyl.

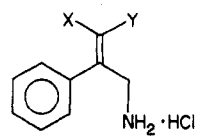
It is now accepted that monoamine oxidase [MAO; monoamine: O_2 oxidoreductase (deaminating), EC 1.4.3.4] exists in two forms with different distribution amongst organs and across species and different sensitivity to inhibitors.¹ The enzyme form that preferentially oxidizes 5-hydroxytryptamine and norepinephrine and is inhibited by low concentrations of clorgyline is designated type A and the form preferentially deaminating phenethylamine and benzylamine and sensitive to low concentrations of L-deprenyl is called type B.²

MAO inhibitors are used in the treatment of depression and certain forms of phobic anxiety and have occasionally been used in the treatment of hypertension.³ Recently, the use of an MAO inhibitor (L-deprenyl) has been found to be beneficial as an adjunct to the L-Dopa treatment of Parkinson's disease.⁴

Over the past three decades several hundred competitive and irreversible inhibitors of MAO have been reported.⁵ The vast majority of these inhibitors show no selectivity for either the A or B form of the enzyme, the major exceptions being the A-selective inhibitors clorgyline, Lilly 51641, harmine, MD 780515, FLA 314, and Ro 11-1163 and the B-selective inhibitors L-deprenyl, pargyline, and Lilly 54761.⁶

Not long after introduction of MAO inhibitors into clinical practice a serious problem associated with their use was discovered.⁷ Severe hypertensive crises can arise in MAO inhibitor treated patients after the ingestion of certain foodstuffs (e.g., cheese) containing the pressor amine tyramine. This so-called "cheese effect" appears to be associated essentially with inhibition of the A form of MAO.⁸ L-Deprenyl, an irreversible, B-selective inhibitor of MAO which has been extensively studied in the clinic,

Table I. Influence of Halogen Substitution on MAO Potency



compd	X	Y	mp, °C	formula ^a	IC_{50} , ^b M
1	H	H	178-179 ^c	$C_9H_{11}N \cdot HCl$	4×10^{-4}
2	F	H	195-196	$C_9H_{10}FN \cdot HCl$	1.8×10^{-8}
3	H	F	144-145	$C_9H_{10}FN \cdot HCl$	1.5×10^{-7}
4	Cl	H	185-186	$C_9H_{10}ClN \cdot HCl$	1×10^{-4}
5	H	Cl	156-157	$C_9H_{10}ClN \cdot HCl$	5×10^{-4}
6	Br	H	190-191	$C_9H_{10}BrN \cdot HCl$	4×10^{-5}
7	F	F	139-140	$C_9H_8F_2N \cdot HCl$	1×10^{-5}
8	Br	Br	243-244	$C_9H_8Br_2N \cdot HCl$	5×10^{-4}
allylamine					5×10^{-4}

^a Satisfactory C, H, N analyses were obtained for compounds 1-8. ^b IC_{50} values were calculated graphically from the inhibition curves obtained when various concentrations (10^{-8} - 10^{-10} M) of the compounds were preincubated with a crude preparation of rat brain mitochondrial MAO for 15 min before measuring the remaining enzyme activity by continuing the incubation for 60 min with tyramine as substrate. ^c Lit.^{13b} mp 140-142 °C.

appears to be free of the cheese effect.⁹ This inhibitor, however, has indirect sympathomimetic activity which is probably due to its rapid metabolism to methamphetamine and amphetamine¹⁰ in addition to its property to inhibit monoamine uptake.¹¹ These secondary effects complicate the interpretation of L-deprenyl's clinical activity solely in terms of inhibition of MAO B. We have recently described¹² the properties of (*E*)-2-(3,4-dimethoxyphenyl)-3-fluoroallylamine (**9**), a selective MAO B inhibitor without indirect sympathomimetic or uptake blocking properties. Since **9** is an interesting member of a new class of MAO inhibitors, we have examined the influence of vinylic halogen substitution and ring substitution of these phenylallylamine derivatives on the potency and/or selectivity for the A and B forms of MAO.

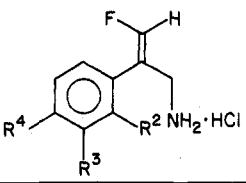
Results and Discussion

Chemistry. The compounds synthesized as a part of this study are listed in Tables I and II. The parent compound **1** has been reported several times¹³ and is readily

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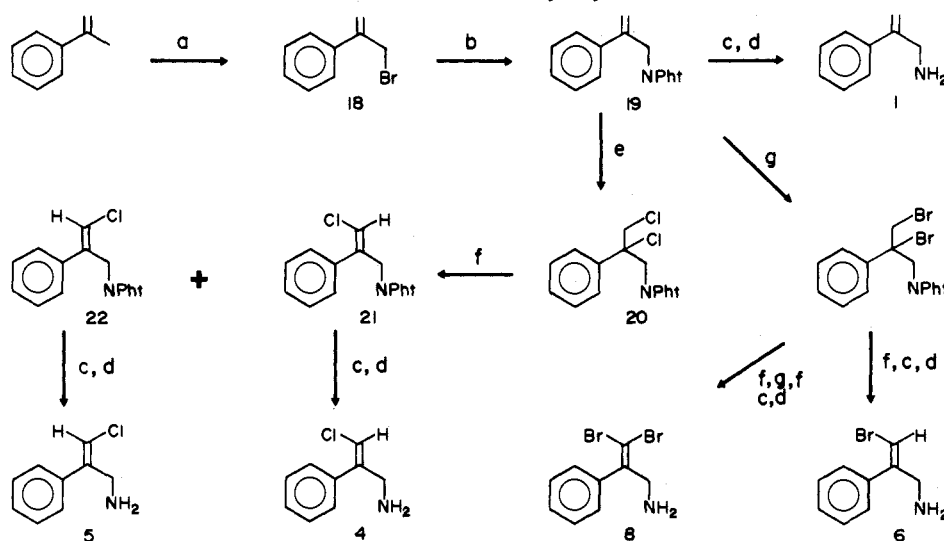
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Table II. Influence of Ring Substitution on MAO Potency



compd	R ²	R ³	R ⁴	mp, °C	formula ^a	IC ₅₀ , ^b M	selectivity ^c
2	H	H	H	195–196	C ₉ H ₁₀ FN·HCl	1.8 × 10 ⁻⁸	10
9	H	OCH ₃	OCH ₃	216–217	C ₁₁ H ₁₄ FNO ₂ ·HCl	2.5 × 10 ⁻⁸	100
10	OCH ₃	H	H	224–225	C ₁₀ H ₁₂ FNO·HCl	2.5 × 10 ⁻⁸	2
11	H	OCH ₃	H	146–147	C ₁₀ H ₁₂ FNO·HCl	5 × 10 ⁻⁹	4
12	H	H	OCH ₃	172–173	C ₁₀ H ₁₂ FNO·HCl	6 × 10 ⁻⁸	100
13	H	H	Cl	189–190	C ₉ H ₉ ClFN·HCl	2 × 10 ⁻⁸	10
14	H	H	CH ₃	169–170	C ₁₀ H ₁₂ FN·HCl	1 × 10 ⁻⁸	5
15	H	CF ₃	H	174–175	C ₁₀ H ₉ F ₄ N·HCl	7 × 10 ⁻⁹	2
16	H	OH	H	142–143	C ₉ H ₁₀ FNO·HCl	5 × 10 ⁻⁸	0.1
17	H	OH	OH	192–193	C ₉ H ₁₀ FNO ₂ ·HCl	1 × 10 ⁻⁷	0.1
clorgyline						5 × 10 ⁻⁵	<0.01
L-deprenyl						2.5 × 10 ⁻⁸	100

^aSatisfactory C, H, N analyses were obtained for the new compounds. ^bIC₅₀ values were calculated graphically from the inhibition curves obtained when various concentrations (10⁻⁸–10⁻¹⁰ M) of the compounds were preincubated with a crude preparation of rat brain mitochondrial MAO for 15 min before measuring the remaining enzyme activity by continuing the incubation for 60 min with tyramine as substrate. ^cThe selectivity for the B form relative to the A form of MAO was estimated from the ratio of the concentrations of the inhibitors needed to decrease the activity of both forms of the enzyme at the same rate.

Scheme I. Synthesis of 3-Bromo and 3-Chloro Derivatives of 2-Phenylallylamine^a

^a Reagents: (a) NBS, (b) potassium phthalimide, (c) NH₂NH₂·H₂O, (d) HCl, (e) Cl₂, (f) DBU, (g) Br₂.

available from α -methylstyrene (Scheme I). Thus, treatment of α -(bromomethyl)styrene¹⁴ (18) with potassium phthalimide in DMF afforded 19 from which 1 could be obtained by hydrazinolysis.^{13a} The phthalimido derivative 19 also served as a key intermediate for the synthesis of the bromo- and chlorovinyl analogues. Chlorination of 19 afforded dichloride 20, which was regioselectively dehydrochlorinated with 1,8-diazabicyclo[5.4.0]undec-7-ene (DBU) in Me₂SO to yield a separable mixture of vinyl chlorides 21 and 22. Deprotection of the pure isomers afforded (*E*)- and (*Z*)-2-phenyl-3-chloroallylamine 4 and 5, respectively. Similarly, bromination of 19, followed by dehydrobromination gave a major product which appeared to be the (*E*)-phthalimido derivative, the deprotection of which afforded (*E*)-2-phenyl-3-bromoallylamine (6). Further bromination of this phthalimido derivative gave a tribromide which was dehydrobrominated

and deprotected to yield 2-phenyl-3,3-dibromoallylamine (8).

The syntheses of compounds 2, 7, and 9 to 15 follow the chemistry shown in Scheme II in which the preparation of (*E*)-2-(3,4-dimethoxyphenyl)-3-fluoroallylamine (9) is described as a particular example. The mixed malonate ester (24) was prepared in a straightforward manner from 23, good yields being obtained when 2 equiv of lithium diisopropylamide (LDA) was used as base. Although the introduction of the difluoromethyl group proceeded well under the previously described conditions,¹⁵ somewhat better yields were obtained when sodium *tert*-butoxide was used as base instead of sodium hydride.¹⁶ Decarboxylative fluoride elimination from 25 was achieved in two steps: trifluoroacetic acid to remove the *tert*-butyl ester to give 26, followed by treatment with 1 equiv of sodium hydroxide in aqueous THF. The resulting acrylate ester 27

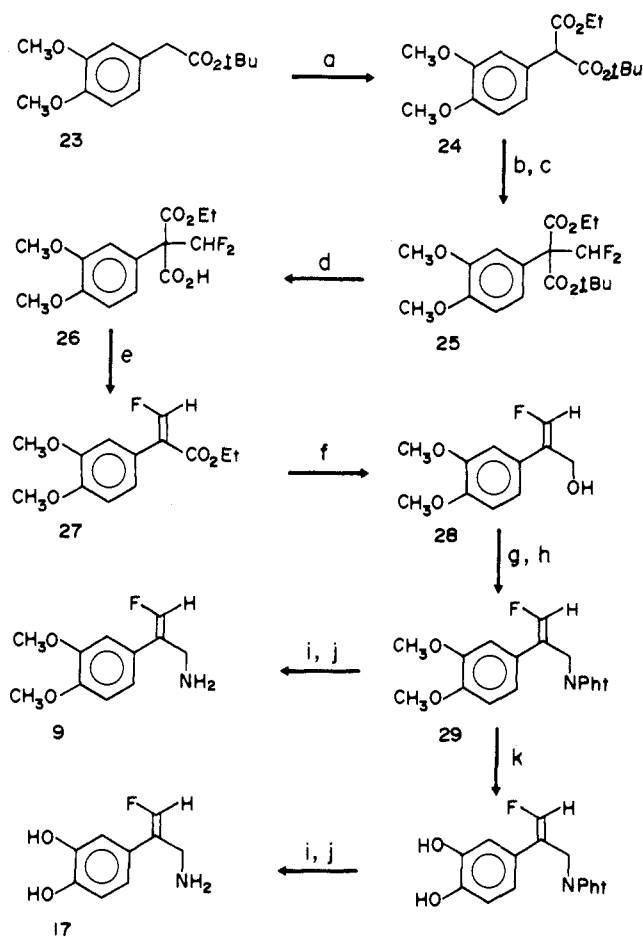
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Scheme II. Synthesis of (*E*)-2-(3,4-Dimethoxyphenyl)-3-fluoroallylamine (9) and (*E*)-2-(3,4-Dihydroxyphenyl)-3-fluoroallylamine (17)^a



^a Reagents: (a) LDA, ClCO_2Et , (b) sodium *tert*-butoxide, (c) Freon 22 (ClCHF_2), (d) $\text{CF}_3\text{CO}_2\text{H}$, (e) NaOH, (f) Dibal, (g) PBr_3 , (h) potassium phthalimide, (i) $\text{NH}_2\text{NH}_2 \cdot \text{H}_2\text{O}$, (j) HCl, (k) BBr_3 .

was selectively reduced with diisobutylaluminum hydride (Dibal) in hexane to give 28 and the phthalimide group was introduced either directly (triphenylphosphine, diethyl azodicarboxylate, phthalimide) or via the bromide (PBr_3 ; then potassium phthalimide in DMF). Deprotection of 29 with hydrazine readily afforded (*E*)-2-(3,4-dimethoxyphenyl)-3-fluoroallylamine (9). In some cases, when the final product did not readily crystallize as its hydrochloride salt, it was found preferable to prepare and purify the *N-tert*-butoxycarbonyl derivative. The amine could then be regenerated as its hydrochloride salt by treatment with hydrogen chloride in anhydrous ether.

A major advantage of this synthesis is that the compounds were obtained exclusively in the *E* configuration. This is due to the stereochemical outcome of the decarboxylative fluoride elimination step in which the (*E*)-acrylate was obtained as the sole isomer. Although this stereoselectivity has been known for many years for decarboxylative chloride or bromide elimination,¹⁷ it was only recently reported for the fluorine case.¹⁸ X-ray structural analyses¹⁹ of (*E*)-2-(4-methoxyphenyl)-3-fluoroallylamine

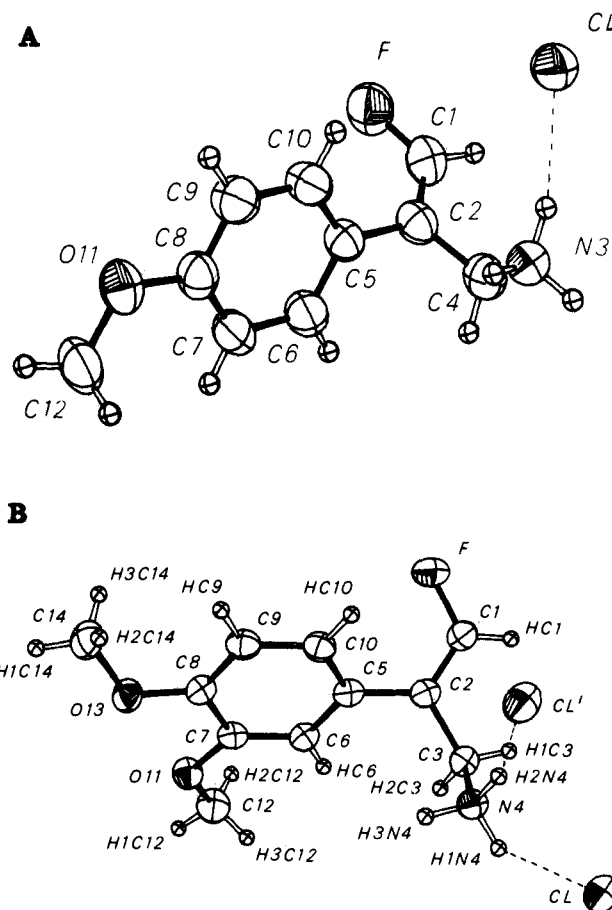
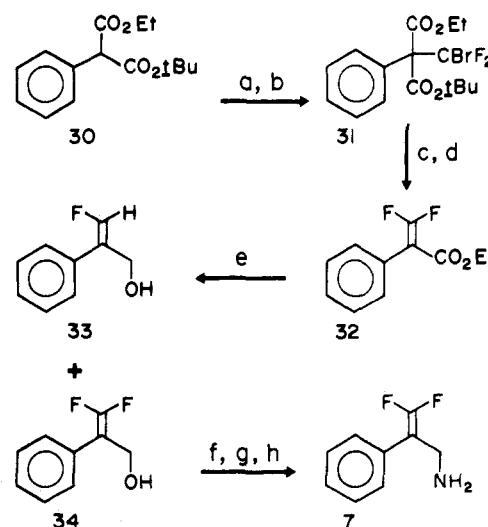


Figure 1. X-ray structural determinations. ORTEP plots of (A) (*E*)-2-(4-methoxyphenyl)-3-fluoroallylamine (12) and (B) (*E*)-2-(3,4-dimethoxyphenyl)-3-fluoroallylamine (9).

Scheme III. Synthesis of 2-Phenyl-3,3-difluoroallylamine (7)^a



^a Reagents: (a) LDA, (b) CBrF_2 , (c) $\text{CF}_3\text{CO}_2\text{H}$, (d) NaOH, (e) Dibal, (f) triphenylphosphine, diethyl azodicarboxylate, phthalimide, (g) $\text{NH}_2\text{NH}_2 \cdot \text{H}_2\text{O}$, (h) HCl.

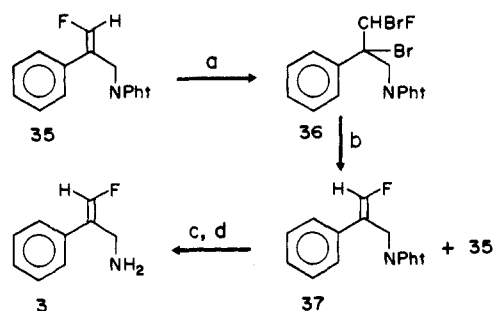
hydrochloride (12, Figure 1A) and (*E*)-2-(3,4-dimethoxyphenyl)-3-fluoroallylamine hydrochloride (9, Figure 1B) confirmed the stereochemistry in the present work.

A complication occurred in the synthesis of 2-phenyl-3,3-difluoroallylamine (7). In distinct contrast with all other cases, Dibal reduction of the acrylate ester 32 gave considerable amounts of (*E*)-2-phenyl-3-fluoroallyl alcohol

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Scheme IV. Synthesis of (*Z*)-2-Phenyl-3-fluoroallylamine (3)^a

^a Reagents: (a) Br₂, (b) NaI, (c) NH₂NH₂·H₂O, (d) HCl.

(33) in addition to the desired 2-phenyl-3,3-difluoroallyl alcohol (34). The ester 32 was obtained in a straightforward manner from the malonate ester 30 with use of dibromodifluoromethane as a means to introduce the bromodifluoromethyl group (Scheme III).

The synthesis of (*Z*)-2-phenyl-3-fluoroallylamine (3) relied upon a *cis* addition of bromine to the double bond in the phthalimide derivative 35, followed by a sodium iodide mediated *trans* debromination (Scheme IV). Although this sequence gave adequate amounts of 3, the initial bromine addition was by no means stereoselective (*cis* and *trans* addition approximately 3 to 1), so that careful chromatography was required to separate phthalimide 37 from 35.

In the syntheses of the *m*-tyramine derivative 16 and dopamine derivative 17 (Scheme II), the methoxyl groups could be cleanly removed with BBr₃ when the nitrogen group was protected as the phthaloyl derivative. Subsequent hydrazinolysis proceeded smoothly to afford the air-stable phenol 16 or catecholamine 17.

Biochemistry. The inhibitory properties of compounds 1–17 toward MAO were initially evaluated in vitro by preincubating a crude preparation of rat brain mitochondrial MAO with various concentrations of inhibitors for 15 min before measuring the remaining MAO activity by continuing the incubation a further 60 min in the presence of the mixed substrate tyramine. The approximate IC₅₀ values, presented in Tables I and II, were calculated graphically from the inhibition curves obtained with inhibitor concentrations from 10⁻³ to 10⁻¹⁰ M. In Table II, the IC₅₀ values of clorgyline and L-deprenyl, determined under the same conditions, are included for comparison purposes. The inhibitors were further assessed as time-dependent inhibitors of MAO by using the previously described method.¹² With the exception of the *Z*-chloro analogue 5, which acted as a competitive inhibitor, all inhibitors caused a time-dependent inhibition of MAO when selective substrates were used. Pseudo-first-order kinetics were usually observed²⁰ for the inactivation of both MAO A and MAO B; i.e., when 5-HT and phenethylamine, respectively, were used as substrates (e.g., for 2 see Figure 2). Selectivity for the A and B forms of the enzyme was approximated by determining for each inhibitor the concentrations necessary to decrease the activity of the A and B forms at the same rate. The concentrations were selected so that 50% loss of enzyme activity occurred within the range of 5–20 min. The ratio of these two concentrations, listed in Table II, was taken as an approximate

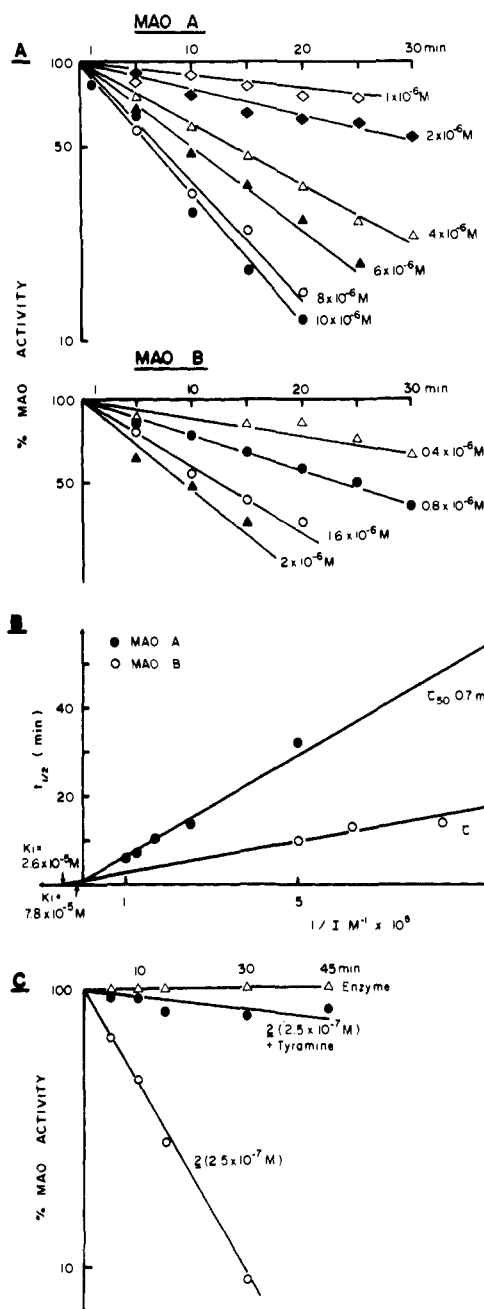


Figure 2. Time-dependent inhibition of MAO by (*E*)-2-phenyl-3-fluoroallylamine (2) at 3 °C.²⁰ (A) Time-dependent inhibition of MAO measured with the selective substrates 5-HT (upper part) and phenethylamine (lower part); (B) plot of $t_{1/2}$ vs. $1/[I]$ according to Kitz and Wilson;³⁰ (C) protection against inactivation by the substrate tyramine (5 mM) at 37 °C.

index of selectivity for the B form relative to the A form of MAO.

With use of (*E*)-2-phenyl-3-fluoroallylamine (2) as a representative example, the mechanism of inhibition was investigated. Protection of MAO against inhibition by 2 occurred in the presence of substrate. When the time course of inactivation by 2 (5×10^{-7} M) was followed in the presence of tyramine (5 mM), complete protection was achieved (Figure 2). The rate of inhibition was not modified by the presence of mercaptoethanol (1 mM) in the incubation medium, indicating that the reactive intermediate generated from 2 was not being released from the active site and then inhibiting the enzyme via an affinity labeling mode. Enzyme inactivation was irreversible as shown by the lack of recovery of enzyme activity upon extensive dialysis of the inactivated enzyme. Taken to-

(20) To analyze the kinetic data for some of these compounds (e.g., 2 and 9) it was necessary to work at lower temperatures (3 or 10 °C) in order to reduce the rate of inactivation so that saturation kinetics were clearly apparent.

gether, these results strongly suggest that the fluoroallylamines derivatives are enzyme-activated inhibitors²¹ of MAO.

Structure-Activity Relationships. We have confirmed Rando's results²² that allylamine is a weak inhibitor of MAO ($IC_{50} = 5 \times 10^{-4}$ M) in vitro. Rando also reported²³ that 3-bromo- and 3-chloroallylamine are MAO inhibitors of similar potencies. Modification of the simple allylamine structure by the introduction of a phenyl group did not lead to increased potency, 2-phenylallylamine (1) being a relatively poor inhibitor of MAO ($IC_{50} = 4 \times 10^{-4}$ M). Similarly, the vinylic chlorine (4 and 5) and bromine (6 and 8) derivatives of 1 were weak inhibitors. In marked contrast, the vinylic monofluorine derivatives 2 and 3 were much more potent. The most active compound of the nonsubstituted aromatic ring series (Table I) was the *E* isomer 2 ($IC_{50} = 1.8 \times 10^{-8}$ M).

It is not clear why vinylic fluorine substitution leads to greatly enhanced inhibitory potency in contrast to bromine or chlorine. If the mechanism of inhibition, as suggested by Rando²² for the inactivation of MAO by bromo- and chloroallylamine, proceeds via addition of an enzyme residue (or the flavin cofactor) to an α,β -unsaturated imine species followed by halogen elimination, fluorine substitution compared to bromine and chlorine could be advantageous. The strong inductive properties of fluorine, coupled with its small size, could act to increase the electrophilicity of the conjugated imine without perturbing the steric environment. The recent availability of radiolabeled 9 should allow us to study the mechanism of inhibition in detail, hopefully leading to an understanding of the importance at fluorine substitution and stereochemistry. The X-ray structural information may be useful in this regard, especially when the *Z* isomer can be studied, a task which is proving difficult due to the relative instability and poor crystal form of this isomer.

Ring substitution in the vinylic fluorine series (Table II; 2 and 9-17) was shown to have an influence on both inhibitory potency as well as on selectivity for the A and B forms of the enzyme. The most potent compound in this series was the 3-methoxyphenyl derivative 11 ($IC_{50} = 5 \times 10^{-9}$ M), being about 10^5 times more potent than 2-phenylallylamine (1). With the exception of the hydroxylated derivatives 16 and 17, the fluoroallylamine derivatives (Table II) showed selectivity for the B form of the enzyme. The most selective compounds were the 4-methoxy (12) and 3,4-dimethoxy (9) derivatives, both being in vitro as selective for the B form of MAO as L-deprenyl (approximately 100-fold). On the other hand, hydroxyl substitution in the 3- and/or 4-position of the aromatic ring led to A selectivity (approximately 10-fold).

These observations are in general agreement with what has been reported in the literature concerning substrate preference for one form of the enzyme or the other. For example, at low substrate concentrations the methoxyl-substituted substances 4-methoxyphenethylamine and, to a lesser extent, 3,4-dimethoxyphenethylamine^{24,25} would appear to be preferentially oxidized by rat brain mitochondrial MAO B while the hydroxy derivatives nor-epinephrine² and to a lesser extent dopamine²⁶ and octa-

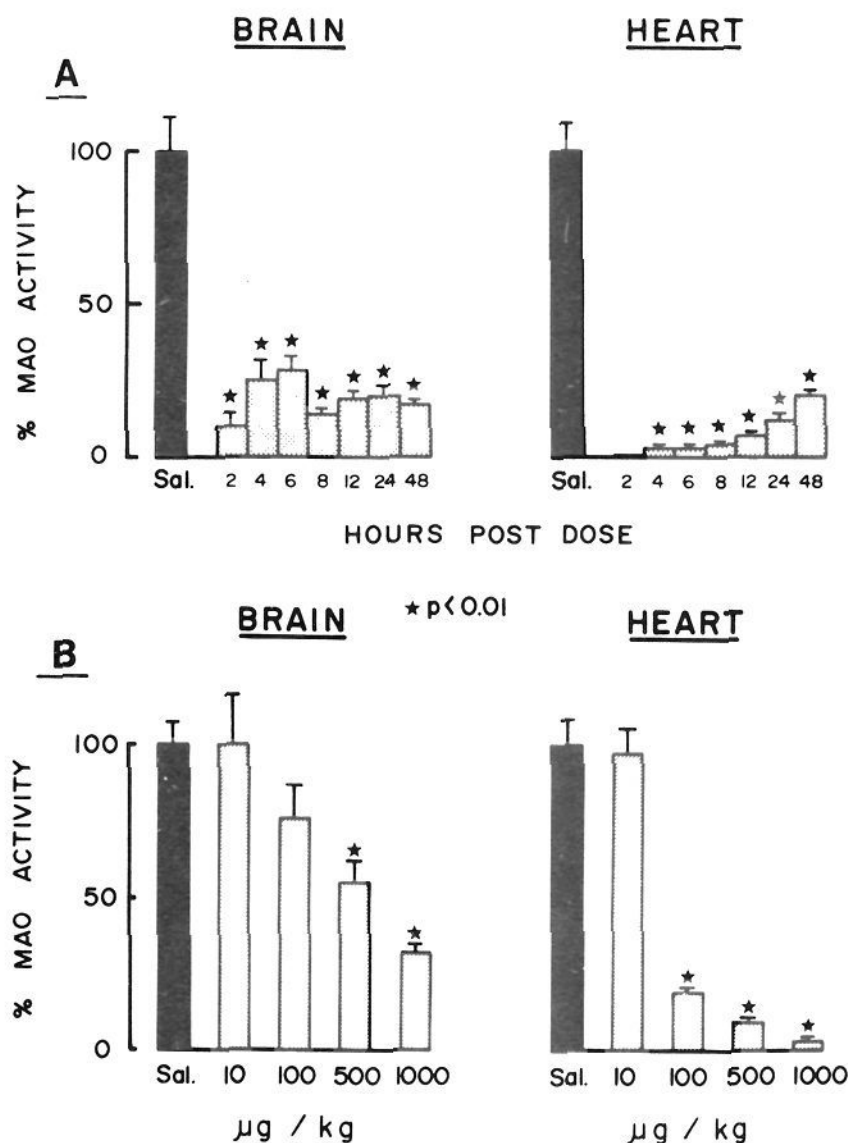


Figure 3. Time course (A) and dose-response (B) of inhibition of MAO in mice ($n = 5$) by (*E*)-2-phenyl-3-fluoroallylamine (2). Mice were injected ip with 1 mg/kg. Residual MAO activity was measured 24 h after the dose.

pamine²⁷ are metabolized mainly by the A form. *p*-Tyramine is considered a mixed substrate. It is conceivable, therefore, that in this series of inhibitors para methoxy substitution leads to B-selective inhibitors and meta hydroxy substitution favors the formation of A-selective inhibitors. Whether this relationship is apparent when the inhibitors are evaluated with MAO from other tissues apart from rat brain has not yet been established. Recently Tipton and Fowler²⁶ emphasized the importance of the tissue source of MAO with respect to substrate specificity. Preliminary results indicate that the nature of vinylic substitution, whether hydrogen, chlorine, or fluorine, has little influence on selectivity. Finally, all other factors being equal, side-chain β -substitution appears to be associated with selectivity for the B form of the enzyme. This is in agreement with the observation that β -phenylethanolamine is a preferential substrate for MAO B and complements the A selectivity often seen with α -methyl-substituted reversible inhibitors.

Pharmacology. Some of these compounds have been examined as MAO inhibitors in vivo. The non-ring-substituted 2, for example, given to mice at 1 mg/kg ip, gave good inhibition in the brain and heart (as well as in other tissues), which lasted for at least 48 h (Figures 3A and 3B). This inhibition was accompanied by the expected increases in noradrenaline, dopamine, and 5-hydroxytryptamine with concomitant reduction of their deaminated metabolites (Figure 4).

The most interesting compound in the series, the B-selective inhibitor 9, has been selected for further development. Preliminary results of current biochemical and

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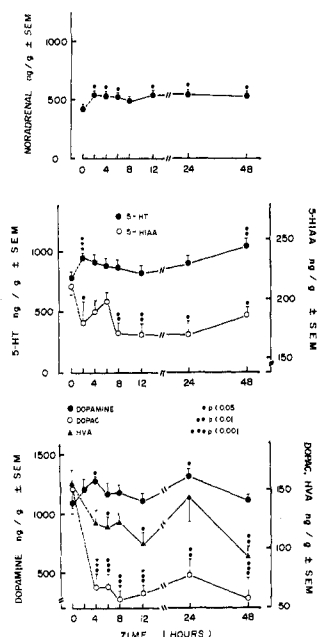


Figure 4. Time course of the effect of a single dose (1 mg/kg ip) of (*E*)-2-phenyl-3-fluoroallylamine (**2**) on mouse brain ($n = 5$) monoamine and monoamine metabolite concentrations. 5-HT, 5-hydroxytryptamine; 5-HIAA, 5-hydroxyindole-3-acetic acid; DOPAC, dihydroxyphenylacetic acid; HVA, homovanillic acid.

pharmacology studies have recently been reported.²⁸

Experimental Section

Chemistry. Melting points were determined on a Mettler FP5 instrument or a Büchi SMP20 instrument and are uncorrected. Microanalyses were obtained on a Perkin-Elmer 240 CHN analyzer. ¹H NMR spectra were recorded on a Varian Associates Model T-60 spectrophotometer using trimethylsilane (organic solutions) or 3-(trimethylsilyl)propionic acid-*d*₄ sodium salt (D₂O solutions) as internal standard.

2-Phenyl-3-phthalimidopropene (19).^{13a} A mixture (30.13 g) of α -(bromomethyl)styrene (0.115 mol) and 1-bromo-2-phenylpropene, prepared according to Reed,¹⁴ and potassium phthalimide (21.20 g, 0.115 mol) in DMF (100 mL) was treated at 90 °C for 3 h. The mixture was cooled, diluted with water, and extracted with CHCl₃. The CHCl₃ extracted was washed with brine, dried, and evaporated to leave a partially solid mass. Trituration with CH₃OH, followed by recrystallization from CHCl₃/petroleum ether afforded **19** (18.6 g) as colorless needles; mp 122–124 °C (lit.^{13a} mp 122 °C).

2-Phenylallylamine (1).^{13a} The hydrochloride salt was recrystallized from EtOH/EtOAc as colorless plates; mp 178–179 °C (lit.^{13b} mp 140–142 °C); NMR (D₂O) δ 4.07 (s, 2 H), 5.43 (m, 1 H), 5.66 (s, 1 H), 7.45 (s, 5 H).

1,2-Dichloro-2-phenyl-3-phthalimidopropene (20). Chlorine gas was bubbled for 15 min through a solution of **19** (7.90 g, 30 mmol) in CH₂Cl₂ (100 mL) cooled to 0–5 °C and protected from light. After a further 10 min, the solution was poured into brine and extracted with pentane. The pentane extract was washed with 2% aqueous NaHCO₃ and then with water, dried, and evaporated to yield a colorless mass (9.30 g). Recrystallization from *n*-hexane/CH₂Cl₂ afforded **20** (6.70 g, 67% yield) as colorless

needles; mp 114–115 °C; NMR (CDCl₃) δ 4.22 (AB, $\nu_A = 4.50$, $\nu_B = 3.94$, $J = 12.5$ Hz, 2 H), 4.32 (s, 2 H), 7.17–7.97 (m, 5 H). Anal. (C₁₇H₁₃ClNO₂) C, H, N.

(E)- and (Z)-1-Chloro-2-phenyl-3-phthalimidopropene (21 and 22, respectively). A solution of **20** (4.00 g, 12 mmol) and DBU (2.74 g, 18 mmol) in Me₂SO (200 mL) was heated at 95 °C for 4 h, and then the mixture was cooled, diluted with cold water (300 mL), and extracted with ether. The ether-soluble material (3.98 g, brown oil) was subjected to silica chromatography (10% EtOAc in petroleum ether), and the resulting products were recrystallized from *n*-hexane/CH₂Cl₂. These were the *E* isomer **21** [0.91 g, 26% yield; mp 106–108 °C; NMR (CDCl₃) δ 4.58 (d, $J = 1$ Hz, 2 H), 6.38 (t, distorted, 1 H), 7.30 (s, 5 H), 7.55–7.88 (m, 4 H); anal. (C₁₇H₁₂ClNO₂) C, H, N]; and the *Z* isomer **22** [0.93 g, 26% yield; mp 133–134 °C; NMR (CDCl₃) δ 4.95 (d, $J = 1.8$ Hz, 2 H), 6.42 (t, distorted, 1 H), 7.32 (s, 5 H), 7.57–7.90 (m, 4 H); anal. (C₁₇H₁₂ClNO₂) C, H, N].

(E)-2-Phenyl-3-chloroallylamine (4). A mixture of **21** (445 mg, 1.5 mmol) and hydrazine hydrate (85 mg, 1.7 mmol) in EtOH (3 mL) was refluxed for 1½ h, cooled, treated with 18% aqueous HCl, and heated (90 °C) for a further 45 min. The mixture was cooled and then filtered and the filtrate was evaporated to dryness. The residue was extracted with EtOH and the EtOH solution was evaporated to leave a colorless solid. Recrystallization from EtOH/Et₂O gave the hydrochloride of **4** (142 mg, 47% yield) as colorless needles; mp 185–186 °C; NMR (D₂O/DCI) δ 4.07 (br s, 2 H), 6.78 (m, 1 H), 7.50 (s, 5 H).

(Z)-2-Phenyl-3-chloroallylamine (5). In a similar manner, **22** (445 mg, 1.5 mmol) was converted to the hydrochloride salt of **5** (152 mg, 50% yield); mp 156–157 °C; NMR (D₂O/DCI) δ 4.32 (s, 2 H), 6.75 (s, 1 H), 7.40 (s, 5 H).

(E)-2-Phenyl-3-bromoallylamine (6). The synthesis of **6** from **19** followed essentially the same procedure. Thus, bromination of **19** was achieved with Br₂ in CCl₄ in 91% yield. Dehydrobromination (DBU; 64% yield) to give 1-bromo-2-phenyl-3-phthalimidopropene followed by deprotection (hydrazine hydrate; 69% yield) afforded **6**; colorless needles from EtOH/Et₂O; mp 190–191 °C; NMR (D₂O) δ 4.03 (br s, 2 H), 6.93 (br s, 1 H), 7.48 (s, 5 H).

2-Phenyl-3,3-dibromoallylamine (8). Bromination of 1-bromo-2-phenyl-3-phthalimidopropene followed by dehydrobromination and deprotection as before afforded **8** as colorless needles; mp 243–244 °C; NMR (CD₃OD) δ 4.12 (br s, 2 H), 7.38 (s, 5 H).

Ethyl 2-(tert-Butoxycarbonyl)-2-(3,4-dimethoxyphenyl)acetate (24). A solution of **23** (8.00 g, 31.7 mmol) in THF (50 mL) was added to LDA (63.7 mmol, prepared from 6.43 g of diisopropylamine) in THF (100 mL) at –78 °C. After 1 h at this temperature a solution of ethyl chloroformate (4.15 g, 38.2 mmol) in THF (50 mL) was added, the cooling bath was removed, and the reaction was stirred overnight. The solution was carefully acidified with 6 N aqueous HCl, then the THF was evaporated, and the residue was dissolved in ether. The ether solution was washed with dilute aqueous HCl and then with water, dried, and evaporated. The residue, an almost colorless oil (10.32 g, 100% yield) was used in the following reaction without further purification. From another run a small portion was purified (silica chromatography; 20% Et₂O in petroleum ether) for analysis to give a colorless oil; bp 115 °C (oven) (0.05 mm); NMR (CCl₄) δ 1.25 (t, $J = 7$ Hz, 3 H), 1.43 (s, 9 H), 3.75, 3.78 (2 s, 6 H), 4.12 (q, $J = 7$ Hz, 2 H), 4.25 (s, 1 H), 6.72 (s, 2 H), 6.85 (s, 1 H). Anal. (C₁₇H₂₄O₆) C, H.

Ethyl 2-(tert-Butoxycarbonyl)-2-(difluoromethyl)-2-(3,4-dimethoxyphenyl)acetate (25). A solution of **24** (10.32 g, 31.9 mmol) in THF (20 mL) was added to a stirred suspension of sodium *tert*-butoxide (6.16 g, 64.2 mmol) in THF (80 mL). After stirring for 15 min, the solution was heated to 40–45 °C and a rapid stream of Freon 22 (ClCHF₂) was introduced. The temperature quickly rose to reflux and the addition was continued until the temperature began to drop (after 5–10 min). Stirring was continued for an additional 30 min, and then the mixture was poured into water and extracted with ether. The ether solution was washed with water, dried, and evaporated to afford essentially pure **25** (10.99 g; 92% yield). Distillation of a small portion gave analytically pure material as a colorless oil; bp 105 °C (oven) (0.05 mm); NMR (CCl₄) δ 1.25 (t, $J = 7$ Hz, 3 H), 1.42 (s, 9 H), 3.73 (s, 6 H), 4.20 (q, $J = 7$ Hz, 2 H), 6.25 (t, $J = 56$ Hz,

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1 H), 6.68 (s, 2 H), 6.78 (br s, 1 H). Anal. (C₁₈H₂₄F₂O₆) C, H.

Ethyl (*E*)-2-(3,4-Dimethoxyphenyl)-3-fluoroacrylate (27). A solution of 25 (61.70 g, 0.165 mol) in trifluoroacetic acid (150 mL) was stirred at room temperature for 1 h, and then the solution was evaporated under reduced pressure until all the reagent had been removed. The residue was dissolved in THF (70 mL) and treated with 2 M aqueous NaOH (83 mL, 0.166 mol) for 15 min at room temperature. Then the solution was poured into water and the product was isolated by ether extraction and purified by chromatography (silica, 20% EtOAc in petroleum ether) to give 27 as a colorless solid (39.70 g, 95% yield). Recrystallization of a portion from petroleum ether gave colorless plates; mp 71–72 °C; NMR (CCl₄) δ 1.27 (t, *J* = 7 Hz, 3 H), 3.75 (s, 6 H), 4.15 (q, *J* = 7 Hz, 2 H), 6.72 (s, 3 H), 7.53 (d, *J* = 82 Hz, 1 H). Anal. (C₁₃H₁₅FO₄) C, H.

(*E*)-2-(3,4-Dimethoxyphenyl)-3-fluoroallyl Alcohol (28). A solution of 27 (30 g, 0.118 mol) in a mixture of dry hexane (170 mL) and dry dichloromethane (40 mL) was cooled to 0 °C. To this was added a 1 M solution of Dibal in hexane (260 mL, 0.26 mol) at such a rate that the temperature did not rise above 10 °C (the temperature was also controlled with an acetone–dry ice bath). When all was added (about 20 min), the solution was stirred at ambient temperature for 1 h and then cooled to about 5 °C. Methanol (130 mL) followed by 6 N hydrochloric acid (83 mL) were added consecutively and the temperature kept below 10 °C with external cooling as before. The organic layer was separated; the aqueous layer was extracted several times with ether, and the combined organic solutions were washed with water, dried, and evaporated to afford essentially pure 28 as a pale yellow solid mass (18.60 g; 74% yield). A small portion was recrystallized from *n*-hexane to give colorless plates; mp 56–57 °C; NMR (CDCl₃) δ 2.60 (br s, 1 H), 3.83 (s, 6 H), 4.25 (d, *J* = 5 Hz, 2 H), 6.78 (d, *J* = 83 Hz, 1 H), 6.68–7.20 (m, 3 H). Anal. (C₁₁H₁₃FO₃) C, H.

(*E*)-1-Fluoro-2-(3,4-dimethoxyphenyl)-3-phthalimido-propene (29). **Method A.** A solution of 28 (25.00 g, 0.119 mol), triphenylphosphine (31.23 g, 0.119 mol), and phthalimide (17.52 g, 0.119 mol) in THF (450 mL) was treated with a solution of diethyl azodicarboxylate (20.74 g, 0.119 mol) in THF (100 mL) and then allowed to react overnight. The solvent was evaporated, and the byproducts were largely removed by recrystallization (of the byproducts) from toluene and then from ether. The residue was purified by silica chromatography (20% EtOAc in petroleum ether) to give a major fraction (20.48 g) which was recrystallized from *n*-hexane/CH₂Cl₂ to afford 29 (16.50 g; 41% yield) as colorless plates; mp 102–103 °C; NMR (CDCl₃) δ 3.80, 3.85 (2 overlapping singlets, 6 H), 4.52 (m, 2 H), 6.32 (s, 1/2 H), 6.68–7.28 (m, 3 H), 7.52–7.88 (m, 4 1/2 H). Anal. (C₁₉H₁₆FNO₄) (C₁₉H₁₆FNO₄) C, H, N.

Method B. A solution of 28 (13.72 g, 65.3 mmol) in dry toluene (200 mL) was cooled to about 10 °C and treated with a solution of phosphorus tribromide (7.59 g, 28.1 mmol) in toluene (200 mL). The cooling bath was removed and the reaction was allowed to continue for 1 1/2 h during which time some tar formed. The supernatant was poured into saturated aqueous K₂CO₃ and the product was isolated by ether extraction in the normal way to give brown crystals (16.0 g). This product (16.0 g) and potassium phthalimide (11.32 g, 61.2 mmol) were heated in dry DMF (130 mL) at 65 °C for 4 h, and then the mixture was poured into water and extracted with ether. The ether solution was washed with water, dried, and evaporated to give a solid mass. Recrystallization from *n*-hexane/CH₂Cl₂ afforded 29 (15.95 g, 72% overall yield) as before.

(*E*)-2-(3,4-Dimethoxyphenyl)-3-fluoroallylamine (9). A mixture of 29 (6.82 g, 20 mmol) and hydrazine hydrate (1.10 g, 22 mmol) in CH₃OH (45 mL) was refluxed for 3 h. Aqueous HCl (18%; 12 mL) was added and refluxing was continued for 30 min, and then the mixture was cooled and filtered. Evaporation of the filtrate left a solid residue which was triturated with CH₃OH and then recrystallized from EtOH/Et₂O to afford 9 (2.56 g, 52% yield) as colorless plates; mp 216–217 °C; NMR (D₂O) δ 3.87 (s, 6 H overlapping 4.00, br d, *J* = 4 Hz, 2 H), 7.10 (br s, 3 H), 7.17 (d, *J* = 82 Hz, 1 H).

(*E*)-2-Aryl-3-fluoroallylamines 2 and 10–15. The syntheses of these compounds from the corresponding *tert*-butyl phenylacetates followed the procedures outlined for the synthesis of 9 from 23. Exceptions to this general procedure were as follows:

(1) deprotonation of ethyl 2-(*tert*-butoxycarbonyl)-2-(2-methoxyphenyl)acetate was achieved with *n*-butyllithium/potassium *tert*-butoxide instead of sodium *tert*-butoxide (in the synthesis of 10); (2) decarboxylate fluoride elimination to prepare ethyl (*E*)-2-[3-(trifluoromethyl)phenyl]-3-fluoroacrylate was achieved with triethylamine (2.5 equiv) in CH₂Cl₂ at room temperature (in the synthesis of 15); and (3) Dibal reduction of 32 (see below).

Ethyl 2-(*tert*-Butoxycarbonyl)-2-(bromodifluoromethyl)phenylacetate (31). Following the procedure described for the preparation of 25, ethyl 2-(*tert*-butoxycarbonyl)phenylacetate (15.84 g, 60 mmol) was converted to 31 (19.69 g, 83% yield after chromatography) with use of dibromodifluoromethane in place of Freon 22; NMR (CCl₄) δ 1.28 (t, *J* = 7 Hz, 3 H), 1.52 (s, 9 H), 4.25 (q, *J* = 7 Hz, 2 H), 7.13–7.55 (m, 5 H). Small peaks corresponding to the difluoromethyl and dibromofluoromethyl analogues were also evident in the spectrum.

Ethyl 2-Phenyl-3,3-difluoroacrylate (32). Following the procedure described for the preparation of 27, 31 (20.95 g, 53.3 mmol) was converted to 32 (10.80 g, 96% yield) which was distilled as a colorless oil; bp 70 °C (oven) (0.05 mm); NMR (CCl₄) δ 1.25 (t, *J* = 7 Hz, 3 H); 4.15 (q, *J* = 7 Hz, 2 H); 7.18 (br s, 5 H). This product was contaminated with a small amount of ethyl 2-phenyl-3-bromo-3-fluoroacrylate.

Dibal Reduction of 32. A solution of 32 (7.13 g, 33.6 mmol) in THF (180 mL) was cooled to about –78 °C and a solution of Dibal in hexane (1.3 M solution, 105 mL, 135.6 mmol) was added. The cooling bath was removed and the temperature was allowed to rise over 45 min, and then the solution was cooled to 0–5 °C and treated slowly with methanol (50 mL) and then with 10% aqueous KOH (13.5 mL). The mixture was then dried (MgSO₄), filtered, and evaporated to yield a yellowish oil (4.60 g). Chromatography (silica, 20% EtOAc in petroleum ether) allowed the separation of two products. The first-eluted substance was 34 (1.94 g, 34% yield, colorless oil); NMR (CCl₄) δ 2.70 (s, 1 H), 4.13–4.43 (m, 2 H), 6.98–7.35 (m, 5 H). The second substance was 33 (1.42 g, 28% yield, colorless oil); NMR (CDCl₃) δ 1.68 (s, 1 H), 4.33 (broadened d, *J* = 5 Hz, 2 H), 6.17 (s, 1/2 H), 7.20–7.63 (m, 5 1/2 H). In addition, a mixture (1.00 g) of these two substances was obtained.

2-Phenyl-3,3-difluoroallylamine (7). Alcohol 34 (1.94 g, 11.4 mmol) was converted to 1,1-difluoro-2-phenyl-3-phthalimido-propene (2.12 g, 62% yield) by method A above. This product recrystallized from hexane as colorless needles; mp 102–103 °C; NMR (CCl₄) δ 4.67 (m, 2 H), 7.28 (br s, 5 H), 7.57–7.88 (m, 4 H). Anal. (C₁₇H₁₁F₂NO₂) C, H, N. Treatment of this substance (0.60 g, 2 mmol) with hydrazine hydrate in the normal way gave almost pure amine 7 (0.41 g). Purification was achieved by conversion to the *N*-Boc-protected derivative; colorless needles; mp 44–45 °C; NMR (CCl₄) δ 1.33 (s, 9 H), 3.93–4.27 (m, 2 H), 4.60 (br s, 1 H), 7.27 (s, 5 H). Anal. (C₁₄H₁₇F₂NO₂) C, H, N. Deprotection of this Boc derivative (0.14 g) was achieved with ether/hydrogen chloride whereupon the hydrochloride salt of 7 (0.07 g, 65% yield) crystallized as colorless needles; mp 139–140 °C; NMR (D₂O) δ 4.10 (broadened s, 2 H), 7.43 (s, 5 H).

(*Z*)-2-Phenyl-3-fluoroallylamine (3). In the dark a solution of bromine (0.35 g, 2.2 mmol) in CH₂Cl₂ (5 mL) was added to a cold (0–5 °C) stirred solution of 35 (0.56 g, 2 mmol) in CH₂Cl₂ (30 mL). The cooling bath was then removed and stirring was continued for 24 h. More CH₂Cl₂ was added and the solution was washed consecutively with aqueous Na₂SO₃ and water, dried, and evaporated to leave a colorless solid. Recrystallization from *n*-hexane/CH₂Cl₂ gave 36 (0.66 g, 75% yield) as colorless needles; mp 163–164 °C; NMR (CDCl₃) δ 4.50 (m, 2 H), 7.83 (d, *J* = 47 Hz, 1 H); 7.07–7.90 (m, 9 H). A mixture of 36 (0.66 g, 1.5 mmol) and NaI (15 g, 0.1 mol) in acetone (100 mL) was refluxed for 4 h, cooled, and decolorized with Na₂SO₃. The acetone was evaporated, and the products were isolated by ether extraction. Chromatography (silica; 10% EtOAc in petroleum ether) of the resulting product (0.36 g) gave two pure substances. The first eluted product was the *Z* isomer 37 (0.25 g; 59% yield); colorless needles from CH₂Cl₂/*n*-hexane; mp 78–79 °C; NMR (CDCl₃) δ 4.78 (dd, *J* = 3 Hz, 1.5 Hz), 2 H), 6.80 (dt, *J* = 82 Hz, 1.5 Hz, 1 H), 7.07–7.87 (m, 9 H). Anal. (C₁₇H₁₂FNO₂) C, H, N. The second product was the *E* isomer 35 (0.08 g, 19% yield); colorless needles from *n*-hexane/CH₂Cl₂; mp 98–99 °C; NMR (CDCl₃) δ 4.55 (m, 2 H), 6.32 (broadened s, 1/2 H), 7.13–7.93 (m, 9 1/2 H). Anal.

(C₁₇H₁₂FNO₂) C, H, N. Deprotection of **37** (0.25 g) with hydrazine hydrate gave **3** (0.19 g); the hydrochloride salt recrystallized from EtOH/Et₂O as colorless needles; mp 144–145 °C; NMR (D₂O) δ 4.20 (broadened d, $J = 3$ Hz, 2 H), 7.13 (d, $J = 82$ Hz, 1 H), 7.50 (m, 5 H).

(*E*)-2-(3,4-Dihydroxyphenyl)-3-fluoroallylamine (**17**). A solution of **29** (3.20 g, 9.4 mmol) in CH₂Cl₂ (50 mL) was cooled to -78 °C and treated with BBr₃ (molar solution in CH₂Cl₂; 31 mL, 31 mmol). After 15 min, the cooling bath was removed and the stirring was continued for 30 min. The solution was poured into water and stirred for 30 min, and then the product was isolated by CH₂Cl₂ extraction as a solid mass. Recrystallization from EtOAc/*n*-hexane gave (*E*)-1-fluoro-2-(3,4-dihydroxyphenyl)-3-phthalimidopropene (2.58 g, 88% yield) as cream needles; mp 184–185 °C; NMR (acetone-*d*₆) δ 4.17 (dd, $J = 3$ Hz, 1.5 Hz, 2 H), 6.10–6.33 (m, 3 H), 6.37 (d, $J = 84$ Hz, 1 H), 7.17 (s, 4 H), 7.37 (s, 2 H). Anal. (C₁₇H₁₂FNO₄) C, H, N. This substance was deprotected in the normal way to afford the catecholamine (**17**) which was purified via its *N*-Boc derivative (mp 145–146 °C) whereupon colorless needles of the hydrochloride salt were obtained; mp 192–193 °C NMR (D₂O) δ 3.92 (d, $J = 3$ Hz, 2 H), 6.97 (m, 3 H), 7.07 (d, $J = 81$ Hz, 1 H).

MAO Inhibition. Determination of IC₅₀ Values. Monoamine oxidase (MAO) determination in vitro: Partially purified mitochondrial MAO was prepared from rat brain by homogenization of tissue in 0.1 M phosphate buffer (pH 7.2; 1:2.5 w/v) followed by centrifugation at 12000g for 10 min at 4 °C. The supernatant was recentrifuged at 40000g for 30 min at 4 °C to obtain a mitochondrial pellet which was resuspended in 0.1 M phosphate buffer (pH 7.2; 2.5 mL/g of rat brain). MAO specific activity in the mitochondrial suspension determined using [¹⁴C]tyramine as substrate was 25 nmol (mg protein)⁻¹ h⁻¹. The inhibitors (10⁻³–10⁻¹⁰ M) were preincubated for 15 min with the mitochondrial preparation in 0.1 M phosphate buffer (pH 7.2; 2 mL) and then [¹⁴C]tyramine (20 μ L; prepared by diluting [¹⁴C]tyramine 5 μ Ci; sp act. 50 mCi/mmol with tyramine (100 mM) in 0.1 M phosphate buffer; pH 7.2 to a final volume of 1 mL) was added and the incubation was continued at 37 °C for a further 60 min. Aqueous HCl (2 M, 300 μ L) was added and the mixture was extracted with toluene (7 mL). Radioactivity in the toluene extract was counted by liquid scintillation spectrometry with Econofluor (N.E.N) as the scintillant. Blanks containing either no enzyme or acid-inactivated enzyme were similarly treated. Molar concentrations of inhibitors producing 50% inhibition (IC₅₀) were determined graphically.

Time-Dependent Inhibition. Time-dependent inhibitory kinetics, characteristic of an enzyme-activated irreversible mechanism of action,²¹ were determined by preincubating for various times with different concentrations of the inhibitors the partially purified mitochondrial preparation. Remaining enzyme activity was then determined following a 200-fold dilution of the enzyme. Incubation in 0.1 M phosphate buffer (pH 7.2, 2 mL) of the substrate (either [¹⁴C]tyramine (5 μ M), [¹⁴C]-5-HT (10 μ M), or [¹⁴C]phenethylamine (5 μ M)) was continued for a further 30 min at 37 °C (1 h with tyramine), and then 2 M aqueous HCl (300

μ L) was added. The products were extracted into either toluene/ethyl acetate (1:1) for the 5-HT and phenethylamine metabolites or toluene for tyramine metabolites. Radioactivity was measured as before.

Further Biochemistry. The inhibitor **2** (5 \times 10⁻⁷ M) was incubated in the presence of tyramine (5 mM), and then time-dependent kinetics were determined with [¹⁴C]tyramine as substrate. Similarly, **2** (5 \times 10⁻⁷ M) was incubated in the presence of mercaptoethanol (1 mM). Partially inhibited enzyme (approximately 80% inhibited) was either dialysed against two changes of 0.1 M phosphate buffer (pH 7.2) for 24 h at 4 °C or washed with water (\times 5). Enzyme activity (using [¹⁴C]tyramine) was measured before and after these treatments by using the procedure described for the determination of IC₅₀ values.

MAO Determinations ex Vivo. Groups of mice (male CD₁ albino) were injected intraperitoneally (ip) with various doses of inhibitor and killed 24 h later. In separate experiments mice were injected ip with a single dose of compound and killed at various times after treatment. MAO activity was determined in homogenates of brain and heart with [¹⁴C]tyramine as substrate.

Monoamine and Monoamine Metabolite Determinations. Brains and hearts from the mice treated with the MAO inhibitors were analyzed for their monoamine and metabolite content by HPLC with electrochemical detection according to the method of Wagner et al.²⁹ Statistical evaluation of MAO activity and monoamine determinations was undertaken by using Student's "t" test in comparison to control animals.

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Registry No. 1, 28144-67-4; 1-HCl, 56132-76-4; 2, 85278-69-9; 2-HCl, 85278-05-3; 3, 93605-75-5; 3-HCl, 85278-84-8; 4, 93605-76-6; 4-HCl, 85278-24-6; 5, 93605-77-7; 5-HCl, 85278-23-5; 6, 93605-79-9; 6-HCl, 93605-78-8; 7, 85278-36-0; 7-HCl, 85278-38-2; 7 *N*-Boc deriv, 85278-37-1; 8, 93605-80-2; 8-HCl, 85278-19-9; 9, 85278-66-6; 9-HCl, 85278-04-2; 10, 85278-70-2; 10-HCl, 85278-07-5; 11, 85278-25-7; 11-HCl, 85278-06-4; 12, 85278-71-3; 12-HCl, 85278-11-1; 13, 85278-75-7; 13-HCl, 85278-08-6; 14, 85278-74-6; 14-HCl, 85278-64-4; 15, 85278-73-5; 15-HCl, 85278-67-7; 16, 85278-68-8; 16-HCl, 85278-55-3; 17, 85278-52-0; 17-HCl, 85278-54-2; 17 *N*-Boc deriv, 85278-53-1; 18, 3360-54-1; 19, 16307-59-8; 20, 85278-20-2; 21, 85278-21-3; 22, 85278-22-4; 23, 13790-42-6; 24, 85277-60-7; 25, 85277-70-9; 27, 85277-77-6; 28, 85277-86-7; 29, 85277-95-8; 31, 85278-30-4; 32, 85278-32-6; 33, 85277-87-8; 34, 85278-33-7; 35, 85277-96-9; 36, 93605-81-3; 37, 85278-44-0; MAO, 9001-66-5; potassium phthalimide, 1074-82-4; 2,3-dibromo-2-phenylphthalimidopropene, 85278-16-6; 1-bromo-2-phenyl-3-phthalimidopropene, 93605-82-4; ethyl chloroformate, 541-41-3; Freon 22, 75-45-6; phthalimide, 85-41-6; dibromodifluoromethane, 75-61-6; 1,1-difluoro-2-phenyl-3-phthalimidopropene, 85278-35-9; (*E*)-1-fluoro-2-(3,4-dihydroxyphenyl)-3-phthalimidopropene, 85278-85-9.

Supplementary Material Available: Physical data for the new phenylallylamine derivatives are recorded (27 pages). Ordering information is given on any current masthead page.